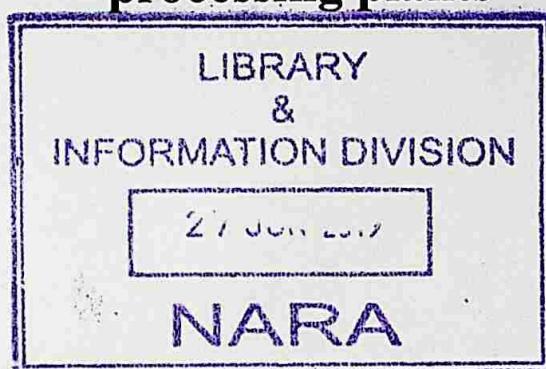


**A study of natural lytic *Listeria* phages with
decontaminating properties for use in seafood
processing plants**



A thesis presented in partial fulfilment of the requirements for the degree of

Doctor of Philosophy

in

Food Technology

Institute of Food, Nutrition and Human Health

Massey University

Auckland, New Zealand

Geevika J Ganegama Arachchi

2013

Table of Contents

Acknowledgements.....	ii
Abstract.....	iv
Table of Contents.....	vi
List of Manuscripts	xi
List of Tables	xii
List of Figures	xiv
Chapter 1: General Introduction	1
References	5
Chapter 2: Literature Review	6
2.1 Introduction.....	6
2.2 Bacteriophage ecology.....	7
2.2.1 Historical background of bacteriophages	7
2.2.2 Existence in nature.....	7
2.2.3 Behaviour of phages and interactions with bacteria	8
2.3 Different applications of phage and phage products	10
2.3.1 Typing phage	10
2.3.2 Use of phages in rapid detection of <i>L. monocytogenes</i> in food	11
2.3.3 Lysins	11
2.3.4 Monocins.....	12
2.4 <i>L. monocytogenes</i> as a food-borne pathogen	13
2.4.1 Prevalence of <i>L. monocytogenes</i>	13
2.4.2 Fish industry and food safety	14
2.4.3 Types of <i>Listeria monocytogenes</i> isolates linked to seafood contaminations	15
2.5 Isolation of phages from environments	17
2.5.1 Isolation of natural lytic phages from the environments	17
2.5.2 Impact of the protocol used for the phage isolation.....	18
2.5.3 Effect of temperature of phage isolation	19
2.6 Characterisation of phage isolates	23
2.6.1 Different parameters to distinguish new phage isolates	23
2.7 Phage application strategies	28
2.7.1 Different approaches of phage application.....	28
2.7.2 Kinetic modelling of phage application	29

2.7.3	Commercial phage products for the food industry	30
2.7.4	<i>Listeria</i> phage-based biocontrol in food systems	31
2.7.5	Phage treatment of abiotic surfaces.....	34
2.7.6	Effectiveness of phage in biofilms	35
2.7.7	Stability of phages.....	36
2.7.8	Development of phage resistant mutants.....	37
2.8	Conclusion	38
	References (Chapter 1 and 2)	39
	Chapter 3: Host range and in vitro lysis of <i>Listeria monocytogenes</i> seafood isolates by bacteriophages	59
	Abstract	60
3.1	Introduction	61
3.2	Materials and methods	63
3.2.1	<i>L. monocytogenes</i> strains and listeriaphages	63
3.2.2	Preparation of phage stocks	63
3.2.3	Host range of phages	64
3.2.4	<i>In vitro</i> lysis of <i>L. monocytogenes</i> by phage FWLLm3	65
3.2.5	Inactivation of low counts of <i>L. monocytogenes</i> by FWLLm3 over 24 h	65
3.2.6	Determination of adsorption rate constant (k value) of phage FWLLm3	65
3.2.7	Estimation of MOI _{input} and minimum phage FWLLm3 titre.....	66
3.2.8	Data analysis	67
3.3	Results and discussion	67
3.3.1	Host range of three phages	67
3.3.2	<i>In vitro</i> lysis of <i>L. monocytogenes</i> cells by phage FWLLm3	72
3.3.3	The k value of phage FWLLm3	75
3.3.4	Estimation of MOI _{input} and minimum dose of phage FWLLm3.....	77
3.3.5	Conclusion.....	78
	Authors' contributions to the manuscript	78
	References	79
	Chapter 4: Characteristics of three listeriaphages isolated from New Zealand seafood environment	85
	Abstract	86
4.1	Introduction	87
4.2	Materials and methods	87
4.2.1	<i>Listeria</i> strains and reference bacteriophage	88
4.2.2	Isolation and purification of phages from environmental samples	88
4.2.3	Plaque morphology at different gel concentrations.....	90

4.2.4	Preparation of purified high titre phage stocks	90
4.2.5	Efficiency of plating (EOP) of phages in citrate agar	90
4.2.6	Heat tolerance of phages at 50 and 60°C	91
4.2.7	Electron microscopy of phages.....	91
4.2.8	Restriction enzyme digestion of phage DNA.....	91
4.2.9	Host range of phages.....	91
4.2.10	Adsorption rate constant (k) of phages	92
4.2.11	Plaque formation temperature range of phages	92
4.2.12	Single-step replication (growth) of phages	93
4.2.13	Statistical analysis.....	93
4.3	Results	93
4.3.1	Isolation of listeriaphages based on plaque characteristics	93
4.3.2	EOP of phages in citrate agar	95
4.3.3	Heat treatment of phages at 50 and 60°C	95
4.3.4	Electron microscope images of phages	96
4.3.5	Phage DNA restriction analysis	98
4.3.6	Host ranges of the phages.....	100
4.3.7	The k values of phages	102
4.3.8	Plaque formation temperature range of phages	103
4.3.9	Single-step replication of phages	103
4.4	Discussion.....	104
4.5	Conclusion	107
	Authors' contributions to the manuscript	108
	References	108
	Supporting Information.....	114
	Chapter 5: Preliminary investigation of bacteriophage-lysis of physiologically stressed <i>L. monocytogenes</i> in seafood processing environments.....	119
	Abstract.....	120
5.1	Introduction.....	121
5.2	Materials and methods	122
5.2.1	<i>L. monocytogenes</i> strains and phages	122
5.2.2	Phage-based lysis of low count of <i>L. monocytogenes</i>	123
5.2.3	Preparation of heat-injured <i>L. monocytogenes</i>	123
5.2.4	Phage-lysis of heat-injured <i>L. monocytogenes</i>	124
5.2.5	Preparation and phage-lysis of starved <i>L. monocytogenes</i>	124
5.2.6	Preparation and phage-based lysis of salt-stressed <i>L. monocytogenes</i>	125

5.2.7	Fluorescence microscopy of stressed <i>L. monocytogenes</i> cells stained with Acridine Orange.....	125
5.2.8	Phage- insensitive cells of high count <i>L. monocytogenes</i> culture.....	125
5.2.9	Determination of N-acetylglucosamine (GLcNAc) in cell wall of <i>L. monocytogenes</i>	126
5.2.10	Statistical analysis	126
5.3	Results and discussion	127
5.3.1	Phage-lysis of low count cells of <i>L. monocytogenes</i> in fish broth	127
5.3.2	Phage-mediated lysis of heat-injured <i>L. monocytogenes</i>	129
5.3.3	Phage-based lysis of starved <i>L. monocytogenes</i> cells	132
5.3.4	Phage-based lysis of salt-stressed <i>L. monocytogenes</i>	133
5.3.5	Phage-insensitive cells of exponential phase <i>L. monocytogenes</i> 19CO9 culture	135
5.3.6	Emergence of phage-resistant <i>L. monocytogenes</i> 19CO9 cells over 48 h	136
5.3.7	Presence of GLcNAc in the cell wall of <i>L. monocytogenes</i>	137
5.3.8	Conclusion.....	139
	Authors' contributions to the manuscript	140
	References	140
Chapter 6: Effectiveness of phages in the decontamination of <i>Listeria monocytogenes</i> adhered to clean stainless steel, stainless steel coated with fish protein, and as a biofilm		145
	Abstract	146
6.1	Introduction	147
6.2	Materials and Methods.....	148
6.2.1	<i>L. monocytogenes</i> strains and <i>Listeria</i> phages.....	148
6.2.2	Test surfaces.....	148
6.2.3	Attachment of <i>L. monocytogenes</i> on surfaces.....	149
6.2.4	Phage-lysis of <i>L. monocytogenes</i> cells adhered on surfaces.....	150
6.2.5	Enumeration of cells adhered to surfaces	150
6.2.6	Preparation of seven-day old <i>L. monocytogenes</i> biofilm on stainless steel coupon	150
6.2.7	Infection of seven-day old biofilm with <i>Listeria</i> phages.....	151
6.2.8	Preparation of dislodged seven-day old biofilm cells	151
6.2.9	Phage-lysis of dislodged seven-day old biofilm cells	152
6.2.10	Fluorescence microscopy of cells on surfaces	152
6.2.11	Data analysis	152
6.3	Results.....	153
6.3.1	Adhesion of <i>L. monocytogenes</i> strains to FBSSC and SSC.....	153

6.3.2	Phage-lysis of <i>L. monocytogenes</i> adhered to surfaces	155
6.3.3	Phage-lysis of seven-day old biofilms on stainless steel.....	156
6.3.4	Phage-lysis of dislodged seven-day old biofilm cells.....	156
6.4	Discussion.....	158
6.5	Conclusion	161
	Authors' contributions to the manuscript	162
	References	162
	Electronic supplementary material (ESM)	169
	Chapter 7: Bacteriophages for the control of low levels of <i>Listeria monocytogenes</i> in a simulated fish processing environment	170
	Abstract.....	171
7.1	Introduction.....	172
7.2	Materials and methods	175
7.2.1	<i>L. monocytogenes</i> strains and <i>Listeria</i> phages.....	175
7.2.2	Preparation of stainless steel coupon (SSC)	175
7.2.3	Fish broth (FB)	176
7.2.4	Phage-based lysis of low count cells on SSC	176
7.2.5	Stability of <i>Listeria</i> phages under different ambient conditions.....	177
7.2.6	Statistical analysis.....	179
7.3	Results	179
7.3.1	Phage-lysis of low counts of <i>L monocytogenes</i> adhered on SSC	179
7.3.2	Stability of <i>Listeria</i> phages under different conditions	181
7.4	Discussion.....	187
7.5	Conclusion	189
	Authors' contributions to the manuscript	190
	References	190
	Highlights.....	195
	Chapter 8: Summary and future directions of study	196
8.1	Summary.....	196
8.2	Future directions of study	198
	Appendix 1	200

Abstract

Listeria monocytogenes is a major cause of illness, associated with seafood, therefore it is important to control this pathogen in seafood processing environments. Sporadic listeriosis outbreaks and seafood recalls indicate that current treatments to control this pathogen may be inadequate. The ability to adapt to harsh environmental conditions, develop resistance and form biofilms makes this environmental pathogen difficult to control using regular disinfectants. Bacteriophages (phages) could serve as effective alternative biocontrol agents. The main objective of this study was to isolate and characterize natural lytic *Listeria* specific phages and examine their effectiveness against *L. monocytogenes* under conditions mimicking those found in seafood processing environments.

Among a group of phages isolated from a seafood waste treatment unit, three phages (LiMN4L, LiMN4p and LiMN17) were selected based on plaque morphology and their source. The three phages were distinguished by morphology, efficiency of plating (EOP) in citrate agar and differences in EOP using different *L. monocytogenes* host strains. Three phages which were found as strictly virulent by whole genome sequence analysis, had broad host ranges at 15 °C and each phage also infected either *L. ivanovii* or *L. innocua*. These phages were unstable at 60 °C for 10 min suggesting psychrotrophic properties. The three phages showed low burst sizes indicating their potential suitability as passive biocontrol agents.

Low counts of *L. monocytogenes* strains (19O9, 19DO3 and 19EO3) in late exponential phase, metabolically injured/stressed by heat and salt, lysed by the three phages at 15 °C in 30 min. The results suggested that the three virulent phages may be good candidates as biocontrol agents against *L. monocytogenes* under conditions commonly found in seafood processing plants.

The phages LiMN4L, LiMN4p and LiMN17, used as single phage or a cocktail of three phages, lysed cells adhered to stainless steel conditioned with soluble fish protein and on clean stainless steel coupons (SSC). The phage cocktail also eradicated low cell counts of about 2 log CFU/cm² adhered to SSC surfaces in the presence of fish proteins at 15 °C in 15 min and no re-growth of cells was observed from phage infected surfaces. This study suggested that a biofilm matrix shielded the bacterial cells from phage infection as three consecutive repeat applications of phages did not efficiently lyse undisturbed biofilm cells. Biofilm cells, once removed from the surface, showed

similar to sensitivity to that of exponential phase planktonic cells. Therefore, disruption of the biofilm structure may be required for effective phage treatments.

Phages suspended in phosphate buffered saline survived refrigeration for at least twelve months and were stable for at least 6 h under likely application conditions such as ambient temperatures and under fluorescence lighting. The three phages, either individually or as a cocktail, showed a high lytic efficacy indicating their potential to serve as bio-decontaminating agents in seafood processing environments.